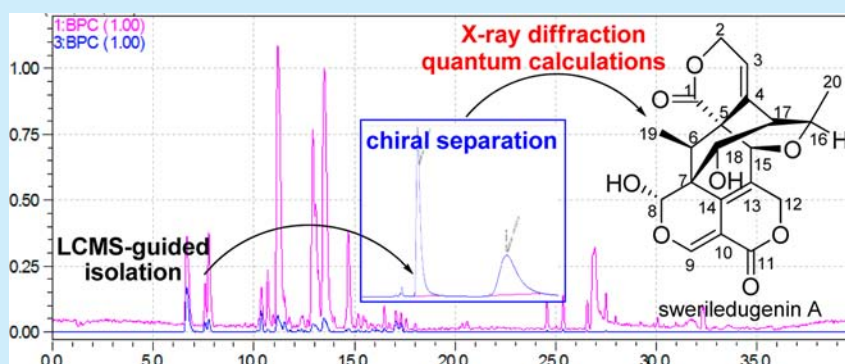


LC-MS Guided Isolation of (\pm)-Sweriledugenin A, a Pair of Enantiomeric Lactones, from *Swertia leducii*

Chang-An Geng, Xing-Long Chen, Ning-Jia Zhou, Hao Chen, Yun-Bao Ma, Xiao-Yan Huang, Xue-Mei Zhang, and Ji-Jun Chen*

State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201, P.R.China

Supporting Information



ABSTRACT: (\pm)-Sweriledugenin A, a pair of novel enantiomeric lactones, were isolated from *Swertia leducii* under the guidance of LC-MS investigation. The enantiomeric separation was achieved by HPLC on a chiral column. Their structures were determined by extensive NMR spectra, X-ray, and quantum calculations. (+)-Sweriledugenin A and (–)-sweriledugenin A showed activities inhibiting HBV DNA replication with the IC_{50} values of 36.86 and 26.55 μ M on the HepG 2.2.15 cell line *in vitro*.

Swertia leducii (Mengzi Zhangyacai in Chinese) is an annual herbaceous plant mainly distributed in Mengzi County of the Yunnan Province. Many traits of *S. leducii* are similar to those of *S. mileensis*, except for petioles and flowers.¹ Therefore, some botanists consider that the two species should be combined. Due to their close morphology, *S. leducii* was always used as the alternative for *S. mileensis* in producing related Qing-Ye-Dan medicines. In our previous investigation, a series of novel secoiridoid dimers and trimers were isolated from *S. mileensis*.² However, no chemical investigation has been conducted on *S. leducii*. Presently, liquid chromatography linked with mass spectrometry (LC-MS) has become a routine method in many areas of analytical chemistry.³ The Shimadzu UFLC-MS-IT-TOF apparatus equipped with an electrospray ionization source coupled to ion-trap and time-of-flight mass analyzers (ESI-IT-TOF) enables high-resolution mass spectra in both positive and negative modes and, thus, is effective for characterizing trace components in a complex mixture of natural products.⁴ To clarify the chemical constituents of *S. leducii*, we used LC-MS guided isolation to obtain a pair of novel enantiomers, (\pm)-sweriledugenin A (**1**), with an unprecedented hexacyclic system (Figure 1). Herein, we reported their isolation, structural elucidation, and anti-HBV activities.

The whole plant of *S. leducii* was collected in Mengzi County, Yunnan Province, China, in October 2012, which was identified as *Swertia leducii* Franch. by Prof. Dr. Li-Gong Lei, Kunming

Institute of Botany, CAS (voucher No. 2012-10-11-1). The dried and powdered whole plants (3.0 kg) were extracted with EtOH (10 L) under reflux 3 times, and the combined solvent was evaporated *in vacuo*. The residue was dissolved in water and partitioned with EtOAc, to afford aqueous and EtOAc parts. The EtOAc part (98 g) was loaded on a silica gel chromatography column (CC) eluting with a MeOH–CHCl₃ system to give six fractions (A–F). Fraction B (5.0 g) was further fractionated by MPLC on an Rp-18 column with MeOH–H₂O as the mobile phase and yielded five subfractions (B1–B5). Subfraction B2 was analyzed by UFLC-MS-IT-TOF to afford a chromatographic peak with the molecular formula C₂₀H₂₀O₈ determined from the [M + H]⁺ ion (m/z 389.1227, –1.03 ppm) in positive mode and the [M – H][–] ion (m/z 387.1084, –0.26 ppm) in negative mode. Consequently, Fr. B2 (100 mg) was purified by HPLC on an Rp-18 column eluted with acetonitrile–H₂O (30:70), and after recrystallization in MeOH, sweriledugenin A (**1**, 3 mg) was obtained.

Sweriledugenin A (**1**)⁵ was isolated as colorless needles with the molecular formula C₂₀H₂₀O₈ which was deduced from [M + H]⁺ (m/z 389.1227) and [M – H][–] (m/z 387.1084) ions in HRESIMS, corresponding to 11 degrees of unsaturation. In

Received: November 6, 2013

Published: December 20, 2013

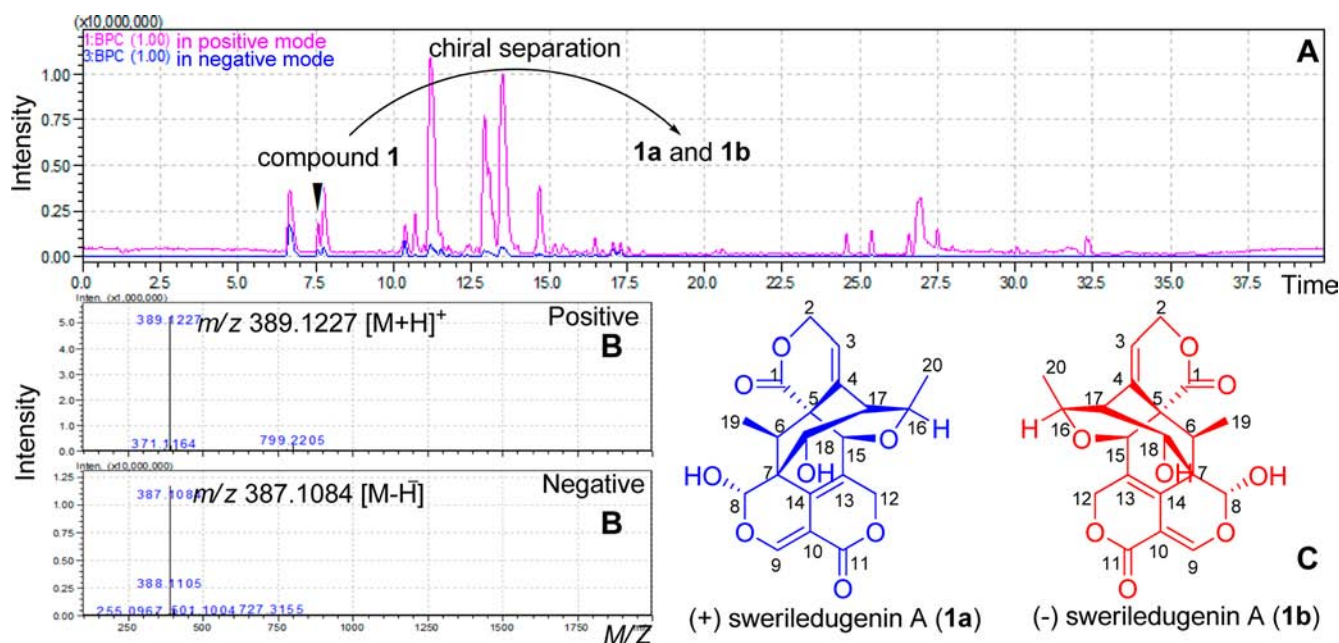


Figure 1. UFLC-MS base peak chromatogram (BPC) of Fr. B2 (A) and HRESIMS (B) as well as structures of **1a** and **1b** (C).

accordance with its molecular formula, all 20 carbons were well resolved in the ^{13}C NMR (DEPT) spectrum, which were recognized as two methyls, two methylenes, eight methines, and eight quaternary carbons. Two ester carbonyl groups (δ_{C} 174.2 and 162.8) and six olefinic carbons (from δ_{C} 151.8 to 104.7) were characterized in the downfield region of ^{13}C NMR spectrum, of which one tetrasubstituted and two trisubstituted double bonds were revealed based on two olefinic protons (δ_{H} 6.03 and 7.39). One doublet at δ_{H} 5.62 (1H, J = 4.6 Hz) in the ^1H NMR spectrum, along with the carbon signal at δ_{C} 97.2 (d), indicated a dioxxygenated methine, which was further proved to be linked with a hydroxyl group by ^1H ^1H COSY of H-8/OH-8. Furthermore, two oxygenated methylenes and two secondary methyls were easily recognized according to the carbons at δ_{C} 70.6 (t), 70.2 (t), 22.5 (q), and 11.4 (q), as well as protons at δ_{H} 1.08 (3H, d, J = 6.3 Hz) and 1.00 (3H, d, J = 7.0 Hz). Based on the above analyses, 5 out of 11 degrees of unsaturation were assigned, and thus, the residual 6 degrees of unsaturation required sweriledugenin A (**1**) to possess a hexacyclic skeleton.

The connectivity of $\text{CH}_3(20)\text{--CH}(16)\text{--CH}(17)\text{--CH}(18)\text{--OH}$ can be well interpreted from the correlations of H-20/H-16/H-17/H-18/OH in the ^1H ^1H COSY spectrum. Similarly, the correlations of H-2/H-3 and H-6/H-19 suggested the direct linkage of $\text{C}(2)\text{--C}(3)$ and $\text{C}(6)\text{--C}(19)$. However, these partial structures could not be assigned with confidence on the basis of the HMBC data. Therefore, an X-ray diffraction analysis was performed, from which the structure of sweriledugenin A was unambiguously determined (Figure 2).⁶ Detailed interpretation of the HMBC (Table 1) and ROESY correlations of H-3/H-17, H-19/H-18 and H-18/H-8 (Figure 3) was well consistent with the structure deduced above.

It is worth noting that the crystal of sweriledugenin A (**1**) had a $p2_1/c$ space group, indicating a racemic nature, which was in accordance with its $[\alpha]_{\text{D}}$ value (+1.76). Subsequent chiral resolution was performed on a chiral column (Daicel Chiralpak AS-H) to yield (+) sweriledugenin A (**1a**) and (–) sweriledugenin A (**1b**), which were opposite in terms of optical rotation (Supporting Information). The final assignment of **1a**

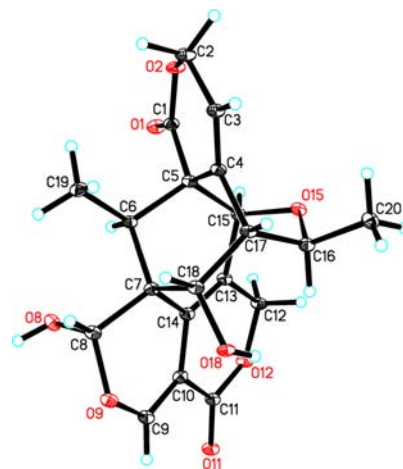


Figure 2. X-ray structure of compound **1**.

and **1b** was achieved using a quantum method by comparing the calculated $[\alpha]_{\text{D}}$ values and electronic circular dichroisms (ECDs) with experimented data.⁷ The absolute configurations of **1a** and **1b** were obtained from X-ray data, and $[\alpha]_{\text{D}}$ values were computed at the b3lyp/6-311g(d,p)//b3lyp/6-311g(d,p) and b3lyp/6-311 +g(2d,p)//b3lyp/6-311+g(2d,p) levels. The calculated $[\alpha]_{\text{D}}$ values were +202/+215 for **1a** and –202/–215 for **1b**, which were very close to the experimented ones of +230 (**1a**) and –172 (**1b**). Similarly, the ECDs calculated at the b3lyp/6-311+g(2d,p) level showed high agreement with the experimented spectra (Figure 4).⁸ From the above evidence, the absolute stereochemistry for **1a** (5*R*, 6*R*, 7*R*, 8*S*, 15*S*, 16*S*, 17*S*, 18*S*) and **1b** (5*S*, 6*S*, 7*S*, 8*R*, 15*R*, 16*R*, 17*R*, 18*R*) were unambiguously determined as shown in Figure 1.

According to the anti-HBV assay on the HepG 2.2.15 cell line *in vitro*,² compounds **1a** and **1b** both showed moderate activity inhibiting HBV DNA replication with the IC_{50} values of 36.86 μM (SI = 10.5) and 26.55 μM (SI = 31.6), respectively.

Sweriledugenin A with a complicated hexacyclic skeleton was isolated under the guidance of the LC-MS method, which was

Table 1. 1D and 2D NMR Data of **1** in Acetone- d_6 (δ in ppm, J in Hz)^a

no.	¹ H NMR ^a	¹³ C NMR ^b	¹ H- ¹ H COSY	HMBC
1	—	174.2, s	—	—
2	5.06, 1H, d, 16.9 4.96, 1H, d, 16.9	70.6, t	H-3	C-1, 4
3	6.03, 1H, t, 2.3	118.1, d	H-2	C-5, 17
4	—	131.1, s	—	—
5	—	49.1, s	—	—
6	2.48, 1H, q, 7.0	45.0, d	H-19	C-1, 4, 8, 14, 18, 19
7	—	44.3, s	—	—
8	5.62, 1H, d, 4.6	97.2, d	HO-8	C-6, 9, 14, 18
9	7.39, 1H, s	151.8, d	—	C-8, 11, 14
10	—	104.7, s	—	—
11	—	162.8, s	—	—
12	4.98, 2H, brs	70.2, t	—	C-11, 14, 15
13	—	118.3, s	—	—
14	—	130.8, s	—	—
15	4.61, 1H, s	75.0, d	—	C-4, 6, 12, 14, 16
16	4.27, 1H, q, 6.3	65.5, d	H-17, H-20	C-4, 15, 18, 20
17	2.57, 1H, d, 5.7	52.5, d	H-16, H-18	C-3, 5, 7, 20
18	4.34, 1H, dd, 5.7, 5.2	73.1, d	H-17	C-4, 6, 8, 14, 16
19	1.00, 3H, d, 7.0	11.4, q	H-6	C-5, 6, 7
20	1.08, 3H, d, 6.3	22.5, q	H-16	C-16, 17
HO-8	6.60, d, 4.6	—	H-8	C-7, 8
HO-18	4.11, 1H, d, 5.2	—	H-18	C-7, 17, 18

^a¹H NMR recorded in 600 MHz. ^b¹³C NMR recorded in 150 MHz.

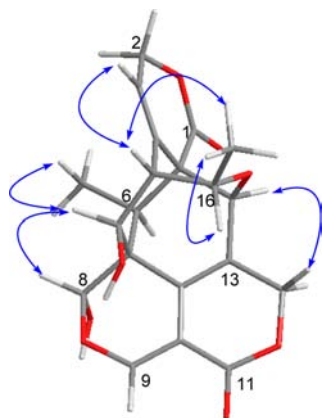


Figure 3. Key ROESY correlations of compound **1**.

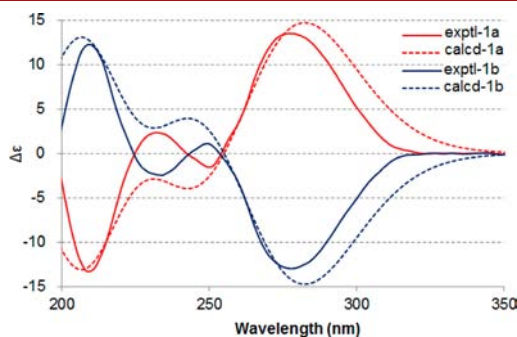


Figure 4. Experimental and calculated ECDs of **1a** and **1b**.

further proven to be a pair of enantiomers by chiral separation and quantum calculations. This investigation is a valuable attempt for guided isolation from a completed natural complex.

■ ASSOCIATED CONTENT

§ Supporting Information

NMR, HRESIMS, $[\alpha]_D$, CD, UV and IR spectra, X-ray data, and computational methods of compound **1**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: chenjj@mail.kib.ac.cn.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

The authors are grateful to Dr. Xiao-Nian Li (Kunming Institute of Botany, CAS) for his kind help in X-ray measurement. The work was supported by the National Science Foundation of China for Distinguished Young Scholars (No. 81025023), the National Natural Science Foundation of China and Yunnan Province (No. U0832603), the West Light Foundation of the Chinese Academy of Sciences, the International Foundation for Science (No. F/5202-1) in Sweden, and the Youth Innovation Promotion Association, CAS.

■ REFERENCES

- (1) *Flora of China* Editorial Committee. *Flora of China*; Science Press: Beijing, 1988; Vol. 62, pp 391–392.
- (2) (a) Geng, C. A.; Jiang, Z. Y.; Ma, Y. B.; Luo, J.; Zhang, X. M.; Wang, H. L.; Shen, Y.; Zuo, A. X.; Zhou, J.; Chen, J. J. *Org. Lett.* **2009**, *11*, 4120–4123. (b) Geng, C. A.; Wang, L. J.; Zhang, X. M.; Ma, Y. B.; Huang, X. Y.; Luo, J.; Guo, R. H.; Zhou, J.; Shen, Y.; Zuo, A. X.; Jiang, Z. Y.; Chen, J. J. *Chem.—Eur. J.* **2011**, *17*, 3893–3903.
- (3) (a) Guan, T. Y.; Liang, Y.; Li, C. Z.; Wang, G. J.; Sheng, L. S. *Chin. J. Nat. Med.* **2011**, *9*, 385–400. (b) Hou, Y. P.; Tianero, M. D. B.; Kwan, J. C.; Wyche, T. P.; Michel, C. R.; Ellis, G. A.; Vazquez-Rivera, E.; Braun, D. R.; Rose, W. E.; Schmidt, E. W.; Bugni, T. S. *Org. Lett.* **2012**, *14*, 5050–5053. (c) Pinto, A.; Wang, M.; Horsman, M.; Boddy, C. N. *Org. Lett.* **2012**, *14*, 2278–2281.
- (4) (a) Xu, X.; Dai, X. J.; Hu, R. L.; Pan, Y. J.; Yang, Z. F. *Molecules* **2011**, *16*, 2551–2560. (b) Geng, C. A.; Ma, Y. B.; Zhang, X. M.; Yao, S. Y.; Xue, D. Q.; Zhang, R. P.; Chen, J. J. *J. Agric. Food Chem.* **2012**, *60*, 8197–8202. (c) Zheng, C. N.; Hao, H. P.; Wang, X.; Wu, X. L.; Wang, G. J.; Sang, G. W.; Liang, Y.; Xie, L.; Xia, C. H.; Yao, X. L. *J. Mass Spectrom.* **2009**, *44*, 230–244. (d) Mori, D.; Kimura, Y.; Kitamura, S.; Sakagami, Y.; Yoshioka, Y.; Shintani, T.; Okamoto, T.; Ojika, M. *J. Org. Chem.* **2007**, *72*, 7190–7198.
- (5) Sweriledugenin A (**1**): colorless crystals (MeOH); mp 266–267 °C; UV λ_{\max} (acetonitrile) (log ϵ) 277 (4.08) nm; IR (KBr) ν_{\max} 3485, 3396, 2929, 1716, 1614, 1456, 1406, 1230, 1096 cm^{-1} ; (+) HRESIMS m/z 389.1227 (calcd for $\text{C}_{20}\text{H}_{21}\text{O}_8$, −0.4 mDa); (−) HRESIMS m/z 387.1084 (calcd for $\text{C}_{20}\text{H}_{19}\text{O}_8$, −0.1 mDa).
- (6) Crystal data for **1**: $\text{C}_{20}\text{H}_{20}\text{O}_8 \cdot \text{CH}_3\text{OH}$, $M = 420.40$, monoclinic, $a = 12.7431(4)$ Å, $b = 11.7643(4)$ Å, $c = 12.0525(4)$ Å, $\alpha = 90.00^\circ$, $\beta = 90.7990(10)^\circ$, $\gamma = 90.00^\circ$, $V = 1806.66(10)$ Å³, $T = 100(2)$ K, space group $P2_1/c$, $Z = 4$, $\mu(\text{Cu K}\alpha) = 1.027$ mm^{-1} , 11 636 reflections measured, 3069 independent reflections ($R_{\text{int}} = 0.0500$). The final R_1 value was 0.1048 ($I > 2\sigma(I)$). The final $wR(F^2)$ value was 0.2894 ($I > 2\sigma(I)$). The final R_1 value was 0.1056 (all data). The final $wR(F^2)$ value was 0.2912 (all data). The goodness of fit on F^2 was 1.445. Crystallographic data have been deposited in the Cambridge Crystallographic Data Centre (deposition number CCDC 957147).

(7) (a) Mazzeo, G.; Santoro, E.; Andolfi, A.; Cimmino, A.; Troselj, P.; Petrovic, A. G.; Superchi, S.; Evidente, A.; Berova, N. *J. Nat. Prod.* **2013**, *76*, 588–599. (b) Zhou, C. X.; Zou, L.; Gan, L. S.; Cao, Y. L. *Org. Lett.* **2013**, *15*, 2734–2737.

(8) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Keith, T.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, O.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. *Gaussian 09*, revision B.01; Gaussian, Inc.: Wallingford, CT, 2010.